

## Enhancement of Relaxivity Rates of Gd–DTPA Complexes by Intercalation into Layered Double Hydroxide Nanoparticles

Zhi Ping Xu,<sup>[a]</sup> Nyoman D. Kurniawan,<sup>[b, c]</sup> Perry F. Bartlett,<sup>[b]</sup> and Gao Qing Lu\*<sup>[a]</sup>

**Abstract:** In this paper we report the preparation and characterization of  $[\text{Gd}(\text{dtpa})]^{2-}$  intercalated layered double hydroxide (LDH) nanomaterials.  $[\text{Gd}(\text{dtpa})]^{2-}$  (gadolinium(III) diethylene triamine pentaacetate) was transferred into LDH by anionic exchange. The intercalation of  $[\text{Gd}(\text{dtpa})]^{2-}$  into LDH was confirmed by X-ray diffraction for the new phase with the interlayer spacing of 3.5–4.0 nm and by FTIR for the characteristic vibration peaks of  $[\text{Gd}(\text{dtpa})]^{2-}$ . The morphology of the nanoparticles was influenced by the extent of  $[\text{Gd}(\text{dtpa})]^{2-}$  loading, in which the poly-dispersity quality decreased as the  $[\text{Gd}(\text{dtpa})]^{2-}$

loading was increased. Compared with the morphology of the original  $\text{Mg}_2\text{Al}-\text{Cl}-\text{LDH}$  nanoparticles (hexagonal plate-like sheets of 50–200 nm), the modified LDH– $\text{Gd}(\text{dtpa})$  nanoparticles are bar-like with a width of 30–60 nm and a length of 50–150 nm. LDH– $\text{Gd}(\text{dtpa})$  was expected to have an increased water proton magnetic resonance relaxivity due to the intercalation of  $[\text{Gd}(\text{dtpa})]^{2-}$  into the LDH interlayer that led to slower

molecular anisotropic tumbling compared with free  $[\text{Gd}(\text{dtpa})]^{2-}$  in solution. Indeed, LDH–nanoparticle suspension containing  $\approx 1.6 \text{ mM}$   $[\text{Gd}(\text{dtpa})]^{2-}$  exhibits a longitudinal proton relaxivity  $r_1$  of  $\approx 16 \text{ mM}^{-1}\text{s}^{-1}$  and a transverse proton relaxivity  $r_2$  of  $\approx 50 \text{ mM}^{-1}\text{s}^{-1}$  at room temperature and a magnetic field of 190 MHz, which represents an enhancement four times ( $r_1$ ) and 12 times ( $r_2$ ) that of free  $[\text{Gd}(\text{dtpa})]^{2-}$  in solution under the same reaction conditions. We have thus tailored LDH–nanoparticles into a novel contrast agent with strong relaxivity, promising for great potential applications in magnetic resonance imaging.

**Keywords:** clays • contrast agents • hydroxides • nanotechnology • proton relaxivity enhancement

### Introduction

Various  $\text{Gd}^{\text{III}}$  complexes have been developed as clinical contrast-enhancing agents in magnetic resonance imaging (MRI), as a tool for clinical diagnosis of organ and tissue

abnormalities.<sup>[1–6]</sup> These complexes enhance the magnetic resonance longitudinal ( $T_1$ ) and transverse ( $T_2$ ) relaxation of the nearby water protons, leading to signal enhancement as observed in MRI. Increasing the level of detection of abnormal organs and tissues in diagnostic MRI is closely linked to the improvement of the water proton relaxivities of these  $\text{Gd}^{\text{III}}$  complexes. A typical example is  $[\text{Gd}(\text{dtpa})(\text{H}_2\text{O})]^{2-}$  (gadolinium(III) diethylene triamine pentaacetate hydrate), as shown in Figure 1. The improvement can be achieved by elongating the rotational correlation lifetime of the nuclear ions<sup>[1,7]</sup> and optimizing the water exchange rate.<sup>[8–14]</sup> Currently, considerable effort is being dedicated to designing new ligand structures, including macrocyclic, polymeric and dendritic frameworks with various functional groups, such as carboxylates, aminos/iminos, phosphates, to increase the rotational correlation lifetime.<sup>[1,6,7]</sup> Some macromolecular ligands have been designed to contain many  $\text{Gd}^{\text{III}}$  atoms and to have a relatively long molecular tumbling time and an optimal exchange rate of the coordinated water molecule (see Figure 1), leading to higher proton relaxivities.<sup>[1,7,9,10]</sup>

Recently, new efforts involving the incorporation of  $\text{Gd}^{\text{III}}$  cations into the framework of inorganic nanoparticles have

[a] Dr. Z. P. Xu, Prof. Dr. G. Q. Lu  
ARC Centre for Functional Nanomaterials  
School of Engineering  
and  
Australia Institute of Bioengineering and Nanotechnology  
The University of Queensland  
Brisbane, QLD 4072 (Australia)  
Fax: (+61) 7-33463973  
E-mail: maxlu@uq.edu.au

[b] Dr. N. D. Kurniawan, Prof. Dr. P. F. Bartlett  
Queensland Brain Institute  
The University of Queensland  
Brisbane, QLD 4072 (Australia)

[c] Dr. N. D. Kurniawan  
Centre for Magnetic Resonance  
The University of Queensland  
Brisbane, QLD 4072 (Australia)

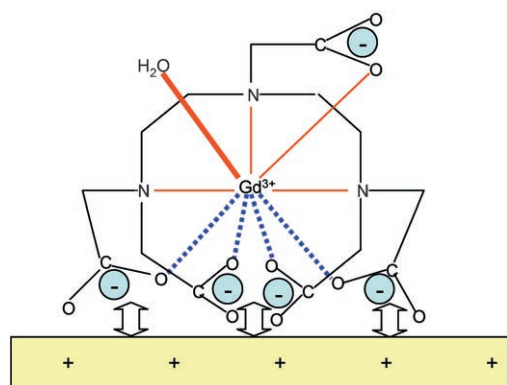


Figure 1. Schematic of possible configuration and interactions between  $[\text{Gd}(\text{dtpa})]^{2-}$  and the positive hydroxide layer.

been reported to improve the relaxivities.<sup>[15–17]</sup> For example,  $\text{Gd}^{\text{III}}$  cations were trapped in the supercage of NaY zeolite<sup>[15]</sup> or incorporated into a mesoporous silica framework.<sup>[16]</sup> In both cases, their magnetic resonance relaxivities increased 3–5 times in the longitudinal axis and 10–20 times in the transverse axis in comparison with those of free  $[\text{Gd}(\text{dtpa})]^{2-}$  ions. In this work, we attempted a new way to enhance the relaxivities of  $\text{Gd}^{\text{III}}$ -complex contrast agents by intercalation into layered double hydroxide (LDH) nanoparticles. This method has indeed enhanced the longitudinal relaxivity by more than four times and the transverse relaxivity by up to 12 times, in relation to the free  $[\text{Gd}(\text{dtpa})]^{2-}$  solution.

LDHs are a group of anion-exchanging materials containing mixed metal hydroxide layer similar to brucite,  $\text{Mg}(\text{OH})_2$ . In a brucite layer, each  $\text{Mg}^{2+}$  ion is octahedrally surrounded by six  $\text{OH}^-$  ions and the different octahedra share edges to form an infinite 2D layer. In hydrotalcite,  $[\text{Mg}_6\text{Al}_2(\text{OH})_{16}][\text{CO}_3 \cdot 4\text{H}_2\text{O}]$ , a quarter of  $\text{Mg}^{2+}$  are replaced by  $\text{Al}^{3+}$ , which gives the brucite-like layers positive charges that are balanced by carbonate anions located in the interlayer region. The positively charge brucite-like layers ( $[\text{Mg}_6\text{Al}_2(\text{OH})_{16}]^{2+}$ ), interlayer anions and water molecules ( $[\text{CO}_3 \cdot 4\text{H}_2\text{O}]^{2-}$ ) are held together via electrostatic interactions and hydrogen bonds to form a 3D structure.<sup>[18,19]</sup> In general, LDHs can be chemically expressed by a formula such as  $[\text{M}^{2+}_{1-x}\text{M}^{3+}_x(\text{OH})_2]^{x+} [(\text{A}^{m-})_{xm} \cdot n\text{H}_2\text{O}]^{x-}$ , where  $\text{M}^{2+}$  represents any divalent cations such as Mg, Zn, Ni, Co, and Fe,  $\text{M}^{3+}$  represents any trivalent cations such as Al, Fe, and Cr, and  $\text{A}^{m-}$  represents the exchangeable anions, such as  $\text{CO}_3^{2-}$ ,  $\text{Cl}^-$ ,  $\text{SO}_4^{2-}$ , various organic anions and complex anions.

$\text{MgAl}$ -LDH possesses physical properties which make it suitable as the carrier of  $[\text{Gd}(\text{dtpa})]^{2-}$  with potential for clinical use: i)  $\text{MgAl}$ -LDH has been shown to be a biocompatible material with low cytotoxicity;<sup>[20]</sup> ii)  $[\text{Gd}(\text{dtpa})]^{2-}$  can be readily intercalated into the LDH interlayer through a chemical exchange process. The chelation of  $\text{Gd}^{\text{III}}$  with DTPA minimizes its toxicity if it de-intercalates from LDH nanoparticles. This is different from the situation in NaY zeolite and mesoporous silica where the  $\text{Gd}^{\text{III}}$  metal ions are

non-chelated, and thus potentially toxic; iii) the  $[\text{Gd}(\text{dtpa})]^{2-}$  intercalated LDH nanoparticles, like the polymeric macromolecules chelating with  $\text{Gd}^{\text{III}}$ , rotate much more slowly, which is expected to promote the relaxations of bound water protons; iv) the particle size and morphology of LDH nanoparticles can be well controlled during the preparation.<sup>[21]</sup> Therefore, in this paper, we intercalated various amounts of  $[\text{Gd}(\text{dtpa})]^{2-}$  [ $\text{Gd}(\text{dtpa})$  hereafter] into  $\text{Mg}_2\text{Al}$ -LDH nanoparticles and investigated the subsequent effect on the water proton relaxivities.

## Results and Discussion

**Intercalation of  $[\text{Gd}(\text{dtpa})]^{2-}$  into the LDH interlayer:** The intercalation of  $[\text{Gd}(\text{dtpa})]^{2-}$  into the  $\text{Mg}_2\text{Al}$ -LDH interlayer is evidenced by the XRD patterns as shown in Figure 2. Obviously, the major phase in each sample is attributed to  $\text{Mg}_2\text{Al}$ -Cl-LDH as the interlayer spacing ranges from 0.766 to 0.773 nm, identical to that reported elsewhere.<sup>[18,19]</sup> However, as marked in Figure 2 (#) for samples E–H, a broad diffraction at around  $2$ – $2.5^\circ$  can be observed. This peak corresponds to an interlayer spacing of 3.5–4.0 nm, which could be attributed to the formation of the  $\text{Mg}_2\text{Al}$ - $\text{Gd}(\text{dtpa})$ -LDH phase. Simultaneously, the diffraction peaks belonging to the  $\text{Mg}_2\text{Al}$ -Cl-LDH phase become broader, as indicated by the increase in FWHM of peak (003) from  $0.34$  to  $0.83^\circ$  (Table 1). In addition, a small bump diffraction at  $18$ – $19^\circ$  (+) in Figure 2 indicates a trace amount of brucite  $[\text{Mg}(\text{OH})_2]$  formed after hydrothermal treatment.<sup>[21]</sup>

The IR spectra illustrated in Figure 3 also indicate the intercalation of  $[\text{Gd}(\text{dtpa})]^{2-}$  into the interlayer. The characteristic IR peaks of  $[\text{Gd}(\text{H}_2\text{dtpa})]$  at  $1591$  and  $1409\text{ cm}^{-1}$  (Figure 3), attributed to the asymmetrical and symmetrical COO stretching vibrations,<sup>[22,23]</sup> gradually intensify from sample A to H at  $1599$  and  $1409\text{ cm}^{-1}$ , respectively. Some finger-printing IR peaks of  $[\text{Gd}(\text{H}_2\text{dtpa})]$  at around  $1000\text{ cm}^{-1}$  can also be distinguished in samples E to H as shoulders. Apart from these, the feature IR peaks of  $\text{Mg}_2\text{Al}$ -LDH can be noted, including a broad band at  $3450\text{ cm}^{-1}$  ( $\nu_{\text{OH}}$ ), a shoulder at  $3000\text{ cm}^{-1}$  ( $\nu_{\text{OH}}$  for O–H bond H-bonded with  $\text{CO}_3^{2-}$ ), a strong peak at  $1365\text{ cm}^{-1}$  [due to  $\nu(\text{CO}_3^{2-})$ ] and bands ( $780$ ,  $683$ ,  $551$  and  $447\text{ cm}^{-1}$ ) below  $1000\text{ cm}^{-1}$  (due to M–O vibrations and M–O–H bending).<sup>[19,24]</sup> In particular, the peak at  $447\text{ cm}^{-1}$  is a unique evidence of 2:1  $\text{MgAl}$ -LDH materials.<sup>[24]</sup> The relatively strong peak at  $1365\text{ cm}^{-1}$  in samples A and C indicates considerable contamination of  $\text{CO}_3^{2-}$  due to the  $\text{CO}_2$  capture during exchanging and washing, which also causes a shoulder at  $3000$ – $3100\text{ cm}^{-1}$ . Note that the shoulder becomes much broader in samples E–H ( $3200$ – $2800\text{ cm}^{-1}$ ), as in  $[\text{Gd}(\text{H}_2\text{dtpa})]$ , due to more H-bonded O–H stretching vibrations that overlap with C–H stretching vibrations.

The elemental analysis shows that more  $[\text{Gd}(\text{dtpa})]^{2-}$  ions are intercalated into the LDH interlayer when more  $[\text{Gd}(\text{dtpa})]^{2-}$  ions are present in the exchange reaction. As can be seen in Table 1, the intercalation degree in LDH nano-

Table 1. Physical characters of Mg<sub>2</sub>Al–Gd(dtpa)/Cl/CO<sub>3</sub>–LDH.

Sample	Molecular formula	2[Gd]/[Al] ratio		FWHM [°]	Average particle size [nm]	PDI	ζ potential [mV]
		Added	Obsd				
A	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.003</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.99</sub> ·nH <sub>2</sub> O	0.01	0.006	0.34	129	0.128	44.2
B	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.012</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.98</sub> ·nH <sub>2</sub> O	0.05	0.024	0.38	133	0.141	46.0
C	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.038</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.92</sub> ·nH <sub>2</sub> O	0.20	0.076	0.60	106	0.100	32.1
D	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.065</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.85</sub> ·nH <sub>2</sub> O	0.40	0.130	0.52	91	0.120	27.2
E	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.099</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.80</sub> ·nH <sub>2</sub> O	0.80	0.198	0.60	83	0.145	25.6
F	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.110</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.78</sub> ·nH <sub>2</sub> O	1.00	0.220	0.76	112	0.266	22.8
G	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.150</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.70</sub> ·nH <sub>2</sub> O	1.50	0.300	0.80	151	0.273	22.1
H	Mg <sup>2+</sup> Al(OH) <sub>6</sub> [Gd(dtpa)] <sub>0.173</sub> (Cl+ <sup>1</sup> / <sub>2</sub> CO <sub>3</sub> ) <sub>0.65</sub> ·nH <sub>2</sub> O	2.00	0.346	0.83	185	0.423	22.0

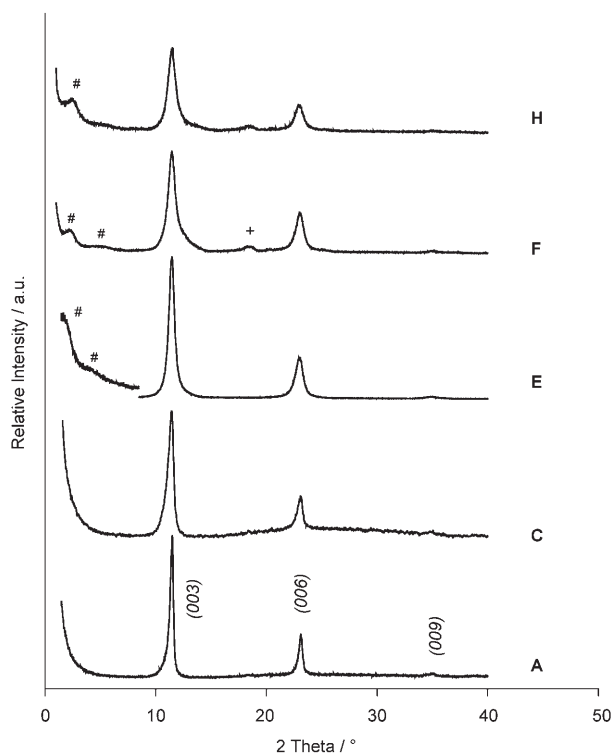


Figure 2. XRD pattern evolution of Mg<sub>2</sub>Al-LDHs with different loadings of [Gd(dtpa)]<sup>2-</sup>. Samples A, C, E, F, H contain 0.05, 0.55, 1.63, 1.71, 2.70 mM [Gd(dtpa)]<sup>2-</sup> (see Table 1).

particles, that is, 2[Gd]/[Al] ratio, gradually increases from 0.006 to 0.346 for samples A to H. However, the intercalation or exchange is relatively incomplete. For example, only about 35% of anions are exchanged even if 200% equivalent [Gd(dtpa)]<sup>2-</sup> are available for the exchange in sample H (Table 1). One possible reason for this incomplete exchange is the contamination of CO<sub>3</sub><sup>2-</sup> that occupy some sites and are difficult to replace. The determinant factor for the lower loading of [Gd(dtpa)]<sup>2-</sup> into Mg<sub>2</sub>Al-LDH, however, is largely a result from its specific size and charge. As estimated from the crystallography data, the dimension of [Gd(dtpa)]<sup>2-</sup> is approximately 1.3 nm.<sup>[25]</sup> Therefore, even with the assumption that [Gd(dtpa)]<sup>2-</sup> packing in the LDH gallery is a bilayer, the closely packed [Gd(dtpa)]<sup>2-</sup> bilayer has only resulted in a negative charge density of 2.4 e nm<sup>-2</sup> (2e × 2/1.3<sup>2</sup> nm<sup>2</sup>), much less than that required to compensate for the positive charges in an Mg<sub>2</sub>Al-hydroxide layer

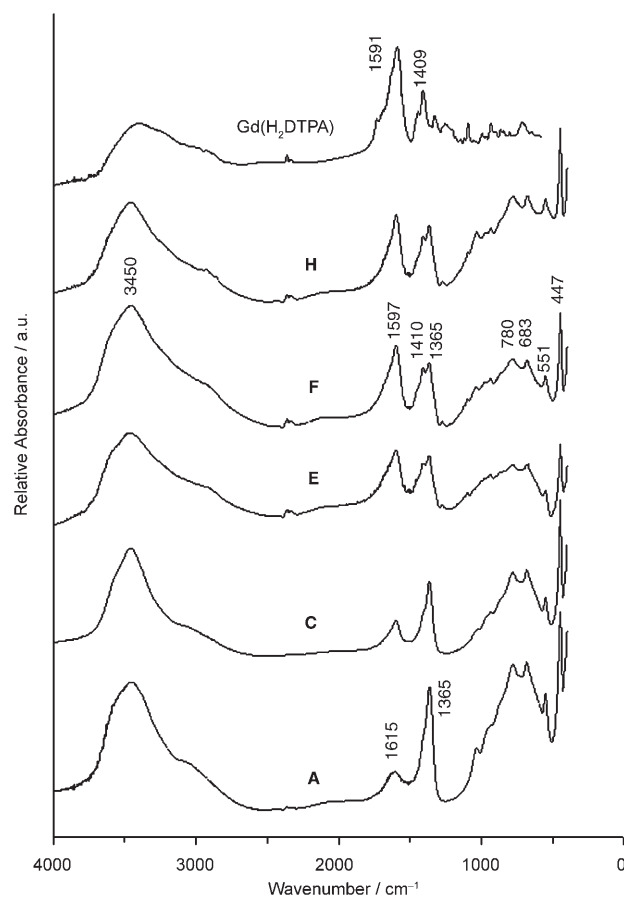


Figure 3. FTIR spectral evolution of Mg<sub>2</sub>Al-LDH with different loadings of [Gd(dtpa)]<sup>2-</sup>.

(4.0 e nm<sup>-2</sup>).<sup>[18]</sup> Therefore about 40% equivalent small anions (such as Cl<sup>-</sup>, CO<sub>3</sub><sup>2-</sup>) must be simultaneously intercalated to compensate for the excess positive charge in the hydroxide layer. This reasoning means that the theoretical exchange degree is 60%, that is, 2[Gd]/[Al] can only reach 0.60 at most. Therefore, the small anions (Cl<sup>-</sup> and CO<sub>3</sub><sup>2-</sup>) have to be accompanied in the sample layer to balance the charges. Seemingly, this Mg<sub>2</sub>Al-[Gd(dtpa)]<sup>2-</sup>/Cl<sup>-</sup>/CO<sub>3</sub><sup>2-</sup>-LDH hybrid phase becomes detectable by XRD when the 2[Gd]/[Al] ratio, that is, the exchange degree, reaches 0.2.

**Size and morphology of Mg<sub>2</sub>Al-[Gd(dtpa)]<sup>2-</sup>-LDH nanoparticles:** After the hydrothermal treatment, the obtained

$[\text{Gd}(\text{dtpa})]^{2-}$ -LDH aqueous suspensions generally appear quite homogeneous. As shown for the particle size distribution in Figure 4, most particles in these samples are in the

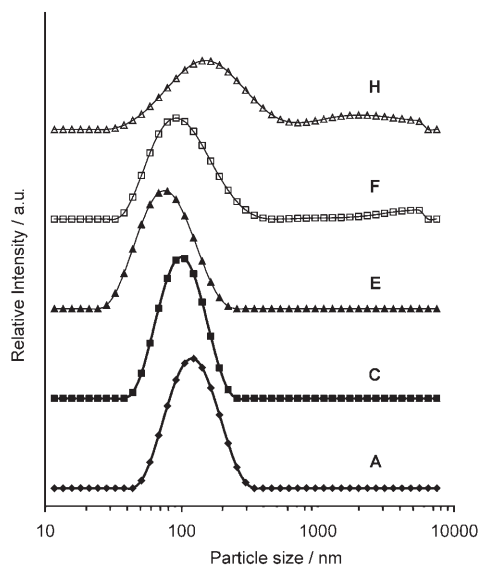


Figure 4. Particle size distribution of as-prepared suspension samples measured with photon correlation spectroscopy (PCS).

range of 40–200 nm. The size distribution is relatively narrow, as indicated by the small polydispersity index (PDI) (from 0.100 to 0.423 in Table 1). Overall, the average hydrodynamic particle size decreases from 129 to 83 nm (sample A to E) and then increases to 185 nm (sample E to H) when more  $[\text{Gd}(\text{dtpa})]^{2-}$  are intercalated. However, the polydispersity of as-prepared Gd(dtpa)-LDH suspensions becomes worse as the PDI increases from 0.100 to 0.423 in the same ordering. In fact, some aggregates in the suspension of sample H were visible. This may be related to the adsorption of some  $[\text{Gd}(\text{dtpa})]^{2-}$  ions on the LDH nanoparticle surface. As shown in Table 1, the zeta potential of these LDH nanoparticle suspensions gradually decreases from 46 to 22 mV for samples A to H, which could be resulted from more  $[\text{Gd}(\text{dtpa})]^{2-}$  anions dynamically located on the surface than  $\text{Cl}^-/\text{CO}_3^{2-}$  ions due to its relatively strong interactions (Figure 1).<sup>[26]</sup> These surface  $[\text{Gd}(\text{dtpa})]^{2-}$  ions have not only led to a low zeta potential, but also caused LDH nanoparticles to aggregate to some degree, as observed in suspension samples F to H.

The morphology of unmodified  $\text{Mg}_2\text{Al}-\text{Cl}/\text{CO}_3$ -LDH nanoparticles prepared by using a similar method is hexagonal plate-like sheets, with a lateral dimension of 50–200 nm, as shown in Figure 5a.<sup>[21,27]</sup> However, in samples F–H, most nanoparticles have a rod-like morphology, while others form irregular thin plates (Figure 5b). In contrast, samples A and B have fewer rod-like nanoparticles but more hexagonal platelets. The nanorods are quite uniform, being 30–60 nm wide and 50–150 nm long, and presumably assigned to the formation of  $\text{Mg}_2\text{Al}-[\text{Gd}(\text{dtpa})]^{2-}/\text{Cl}^-/\text{CO}_3^{2-}$ -LDH hybrid crystals.

Under normal conditions,  $\text{Mg}_2\text{Al}-\text{Cl}/\text{CO}_3$ -LDH crystallites grow preferentially along the *a* and *b* axes, resulting in hexagonal platelets.<sup>[21,27]</sup> However, this growth preference could be altered in the presence of bulky  $[\text{Gd}(\text{dtpa})]^{2-}$  anions. Instead,  $[\text{Gd}(\text{dtpa})]^{2-}$  complex anions probably direct the growth along the *c* axis and thus induce the formation of the nanorod crystals, as in the case of dodecylbenzene sulfonate in  $\text{Mg}_2\text{Al}$ -LDH.<sup>[28]</sup>

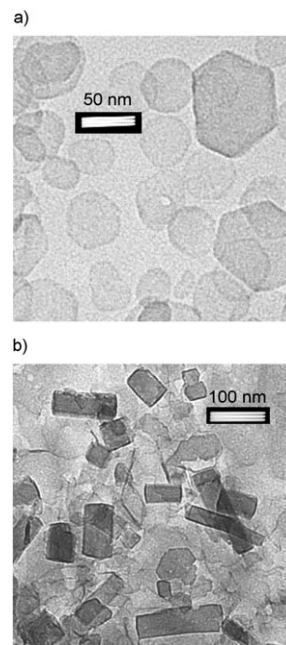


Figure 5. TEM images of typical  $\text{Mg}_2\text{Al}-\text{Cl}/\text{CO}_3$ -LDH (a) and  $\text{Mg}_2\text{Al}-[\text{Gd}(\text{dtpa})]^{2-}/\text{Cl}^-/\text{CO}_3^{2-}$ -LDH nanoparticles (b, sample F).

**Proton relaxivities  $r_1$  and  $r_2$ :** The longitudinal and transverse relaxation times,  $T_1$  and  $T_2$ , were measured for all samples that were freshly prepared in stable suspensions, as described in the Experimental section. The  $T_1$  and  $T_2$  times of all samples and the proton relaxivities,  $r_1$  and  $r_2$  of samples E to H are listed in Table 2. Figure 6 presents the typical

Table 2. Proton relaxation parameters of  $\text{Mg}_2\text{Al}-\text{Gd}(\text{dtpa})/\text{Cl}^-/\text{CO}_3^{2-}$ -LDHs at room temperature and a magnetic field of 190 MHz.

Sample	[Gd] [mM]	$T_1$ [ms]	$T_2$ [ms]	$r_1$ [ $\text{mM}^{-1}\text{s}^{-1}$ ]	$r_2$ [ $\text{mM}^{-1}\text{s}^{-1}$ ]
A	0.05	743	68.2		
B	0.19	298	72.3		
C	0.55	96	29.8		
D	0.97	76	19.7		
E	1.63	36	13.1	16.1	50.0
F	1.71	34	12.4	15.7	47.5
G	2.33	45	10.5	9.5	47.1
H	2.70	54	8.8	6.4	44.8

plots of  $1/T_1$  and  $1/T_2$  versus Gd(dtpa) millimolarity in a series of diluted suspensions, using samples F (■) and H (□) as examples, to determine the relaxivities  $r_1$  and  $r_2$ . Note

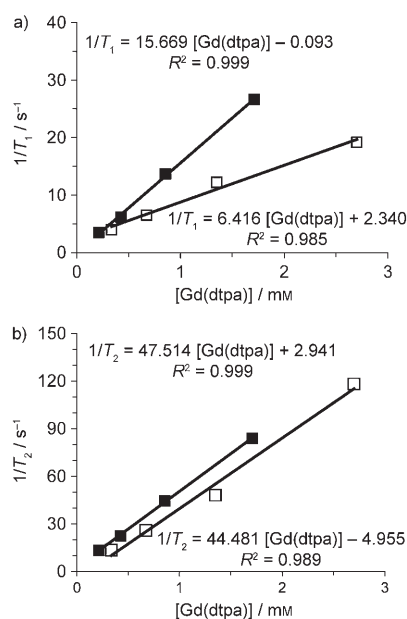


Figure 6. Proton relaxivities  $r_1$  and  $r_2$  of samples F (■) and H (□).

that the longitudinal relaxivity  $r_1$  (6.4–16.1  $\text{mM}^{-1}\text{s}^{-1}$ ) and the transverse relaxivity  $r_2$  (44.8–50.0  $\text{mM}^{-1}\text{s}^{-1}$ ) of these samples are much larger than those of  $[\text{Gd(dtpa)}]^{2-}$  in free solution (3.4 and 4.2  $\text{mM}^{-1}\text{s}^{-1}$  at room temperature and 190 MHz), respectively. It is also noted that when the  $[\text{Gd(dtpa)}]^{2-}$  loading in LDH is increased over 1.7 mM, the  $r_1$  value significantly decreases from 16 to about 6  $\text{mM}^{-1}\text{s}^{-1}$  (Table 2). Consistent with this observation, a more obvious relationship in the current loading range (0.05 to 2.70 mM, Table 2) is presented in Figure 7 for all samples. When  $1/T_1$  and  $1/T_2$  of all samples are plotted against Gd(dtpa) concentration, a linear

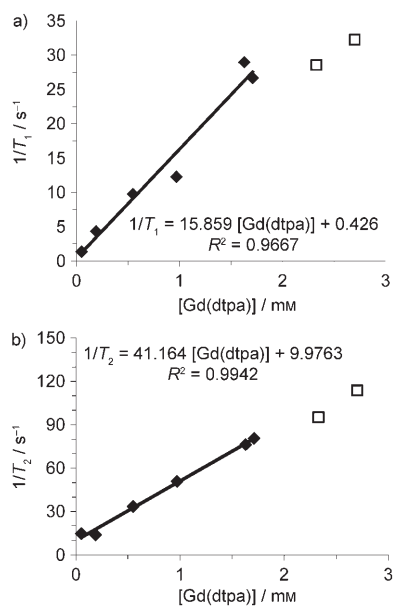


Figure 7. Proton relaxivities  $r_1$  and  $r_2$  of  $\text{Mg}_2\text{Al}[\text{Gd(dtpa)}]^{2-}$ -LDH (for all samples).

relationship is obtained at  $[\text{Gd(dtpa)}]$  below 1.7 mM. The slopes, that is, the average relaxivities  $r_1$  and  $r_2$  of these samples are 15.8 and 41.2  $\text{mM}^{-1}\text{s}^{-1}$ , respectively, identical to those of samples E and F if experimental measurement errors and data treatment errors are considered. These analyses indicate that the proton relaxivities can be enhanced more than four times in the longitudinal axis and 10–12 times in the transverse axis by intercalating  $[\text{Gd(dtpa)}]^{2-}$  ions into the LDH interlayer, as compared with those in free solution. At  $[\text{Gd(dtpa)}]$  above 1.7 mM (Gd wt% in LDH  $\approx 5\%$ ), a decrease in relaxivities  $r_1$  and  $r_2$  is noticeable. A similar decrease in relaxivities was also reported for  $\text{Gd}^{\text{III}}$ -doped mesoporous silica at Gd loading over 2.3% in w/w.<sup>[16]</sup> The decrease in relaxivity is presumably a result of the increase in dipole–dipole interaction of the adjacent nuclear  $\text{Gd}^{\text{III}}$  species that shortens the electronic relaxation, that is, shortens the proton relaxivities<sup>[15,16]</sup> with the decrease of water molecules available for each  $[\text{Gd(dtpa)}]^{2-}$  in the interlayer being another relevant factor, as addressed in the following section.

**Enhancement mechanism:** The relaxivity enhancement of  $\text{Gd}^{\text{III}}$  chelated with macromolecules or incorporated into nanoparticles has been discussed in detail elsewhere.<sup>[1,7]</sup> The major factors for this enhancement include: i) elongation of the rotational correlation lifetime of  $\text{Gd}^{\text{III}}$  species; ii) suitable water exchange rate; and iii) small electron–nuclear spin distance. In the case of Gd(dtpa)-intercalated LDHs, these factors have presumably affected the relaxivities to some degree, in comparison with the free  $[\text{Gd(dtpa)}]^{2-}$  ions.

Obviously, most  $[\text{Gd(dtpa)}]^{2-}$  ions are confined in the LDH interlayer and thus the ionic motions (e.g. rotational and transitional) within the interlayer are much more restricted, in particular,  $[\text{Gd(dtpa)}]^{2-}$  ions are supposed to anchor onto the hydroxide layer through the electrostatic attraction and hydrogen bonds, as schematically drawn in Figure 1. Moreover, the LDH–Gd(dtpa) nanorods (Figure 5b), 30–60 nm wide and 50–150 nm long, tumble much more slowly than the free  $[\text{Gd(dtpa)}]^{2-}$  ions.<sup>[11]</sup> The restricted movement of  $[\text{Gd(dtpa)}]^{2-}$  in the interlayer and very slow nanoparticle tumbling are expected to lead to a longer rotational correlation time, thus increasing the relaxivities.

The exchange rate of water protons in  $[\text{Gd(dtpa)}]^{2-}$ , including the water exchange rate and the prototropic exchange rate,<sup>[8]</sup> may have negative contribution to enhance the relaxivities. For example, the number of water molecules ( $n$ ) in Table 1 is about 1.5 for all samples and thus the available water molecule number per  $[\text{Gd(dtpa)}]^{2-}$  in the interlayer decreased from 500 to 8 for samples A to H. Therefore, the small water number per  $[\text{Gd(dtpa)}]^{2-}$  in samples G and H (8–10) may decrease  $r_1$  relaxivity.<sup>[15]</sup> In zeolite nanoparticles, a typical number of 15–100 water molecules per Gd ion seems to be sufficient to provide a similar level of water exchange rate as in free solution.<sup>[15]</sup>

The main interaction holding  $[\text{Gd(dtpa)}]^{2-}$  anions within the interlayer is the electrostatic attraction between the anions and the positively charged hydroxide layers,<sup>[18]</sup> to-

gether with hydrogen bonds between them. As schematically shown in Figure 1, the attractions would pull the coordinated O atoms (in COO<sup>-</sup> groups) away from the nuclear centre (Gd<sup>3+</sup>) and thus weaken these Gd<sup>3+</sup>...O coordination bonds (dashed blue lines in Figure 1). This bond weakening could be compensated for by the strengthening of the CO bond in the same COO<sup>-</sup> group, as evidenced by the blue shift of the characteristic IR peak from 1591 cm<sup>-1</sup> in [Gd(H<sub>2</sub>dtpa)] to 1599 cm<sup>-1</sup> in LDH (asymmetrical COO<sup>-</sup> stretching vibration). As another consequence, the Gd<sup>3+</sup>...OH<sub>2</sub> bond (bold red line) as well as other coordination bonds (thin red lines) would be strengthened. The strengthening of the Gd<sup>3+</sup>...OH<sub>2</sub> bond, that is, the shortening of the electron-nuclear spin distance, would amplify the proton relaxivities about six times.<sup>[1]</sup>

## Conclusion

In conclusion, [Gd(dtpa)]<sup>2-</sup> has been partially intercalated into Mg<sub>2</sub>Al-LDH nanoparticles by anionic exchange and hydrothermal treatment. The interlayer spacing (3.5–4.0 nm) obtained from XRD data suggests that [Gd(dtpa)]<sup>2-</sup> anions are packed in a bilayer model in the LDH interlayer. Such close packing of [Gd(dtpa)]<sup>2-</sup> still needs some small anions (Cl<sup>-</sup> and CO<sub>3</sub><sup>2-</sup>) together to compensate for the positive charges in the hydroxide layers due to the relatively large size of [Gd(dtpa)]<sup>2-</sup>. The Mg<sub>2</sub>Al-[Gd(dtpa)]<sup>2-</sup>/Cl<sup>-</sup>/CO<sub>3</sub><sup>2-</sup>-LDH hybrid nanocrystals are morphologically rod-like, 30–60 nm wide and 50–150 nm long. Intercalation of [Gd(dtpa)]<sup>2-</sup> into the LDH interlayer enhances the proton relaxivities more than 4 times in the longitudinal axis and up to 12 times in the transverse axis, although high loading of [Gd(dtpa)]<sup>2-</sup> results in relatively small relaxivities. It is found that the major contribution to this enhancement comes from the increase in the rotational correlation lifetime of nuclear species while the shortening of the electron-nuclear spin distance may also contribute to a minor extent.

## Experimental Section

**Preparation of [Gd(dtpa)]<sup>2-</sup>-containing layered double hydroxide nanoparticles:** Layered double hydroxides (LDHs) were prepared by co-precipitation, anion exchange and hydrothermal treatment. In brief, the precursor [Mg<sub>2</sub>Al(OH)<sub>6</sub>Cl]·xH<sub>2</sub>O was prepared by quickly adding (within 5 s) a salt solution mixture (10 mL) containing MgCl<sub>2</sub> (3.0 mmol) and AlCl<sub>3</sub> (1.0 mmol) into a NaOH solution (40 mL, 0.15 M, just enough to form Mg<sub>2</sub>Al-LDH) to co-precipitate under vigorous stirring, followed by 10-minute stirring within a reactor isolated from air. The [Mg<sub>2</sub>Al(OH)<sub>6</sub>Cl]·xH<sub>2</sub>O slurry was obtained via a centrifuge separation and then manually dispersed in a [Na<sub>2</sub>Gd(dtpa)] solution (40 mL) at a designated concentration for the anionic exchange (Table 1). The exchange was allowed to continue for 1 h under vigorous stirring at room temperature. After separating and washing once with deionized water, the Gd(dtpa)-containing LDH slurry was again dispersed in deionized water (40 mL), followed by hydrothermal treatment in an autoclave at 100°C for 16 h, giving rise to a stable homogeneous LDH suspension in most cases. As-prepared suspensions generally have a pH of 8.0 and an Mg<sub>2</sub>Al-Gd(dtpa)/Cl/CO<sub>3</sub>-LDH concentration of 0.4–0.5% in w/w as the

LDH yield of 60 ± 5% has been calculated from the inductively coupled plasma (ICP) data as well as the LDH mass collected from suspensions. In particular, Gd as well as Mg and Al concentrations in the suspensions were determined by the ICP (Tables 1 and 2).

**Characterization of Gd-DTPA-LDH materials:** X-ray diffraction patterns were collected on a glass plate containing a thin layer of LDH hybrid materials at a scanning rate of 1.2° per minute with an interval of 0.02° in each step from 2θ = 1° to 2θ = 40° with CuK<sub>α</sub> radiation (λ = 0.15418 nm) on a Bruker D8 Advance Research X-ray diffractometer by software Diffrac<sup>plus</sup> Evaluation Package Version 11.0. The X-ray diffraction intensity varied within few thousand cps. Infrared spectra were collected on a Perkin-Elmer 2000 FTIR after 40 scans within 4000–400 cm<sup>-1</sup> at a resolution of 4 cm<sup>-1</sup> by measuring the IR absorbance of KBr disc that contains 1–2 wt% of sample. Photon correlation spectroscopy (PCS, Nanosizer Nano ZS, MALVERN Instruments) was used to analyze the particle size distribution of LDH suspensions from 0.6 nm to 6000 nm, in which the peak position (regarded as the average hydrodynamic particle size in the paper) and the polydispersity index (PDI) were automatically generated. The transmission electron microscopic images were obtained on a JEOL JSM-2010 transmission electron microscope (TEM) at an acceleration voltage of 200 kV. For TEM imaging, the freshly collected LDH nanoparticles were dispersed in alcohol with ultrasonication for 20 min, after which a droplet was placed on a copper grid coated with carbon film. In addition, the metal contents of suspension samples were determined by inductively coupled plasma and atomic emission spectroscopy (ICP-AES) on a Varian Vista Pro.

**Water proton relaxivity measurement:** The longitudinal and transverse relaxation times, T<sub>1</sub> and T<sub>2</sub>, of water protons in the as-prepared Gd(dtpa)-LDH suspensions were measured at 190 MHz with a Bruker Biospect 4.6T/20 MR spectrometer at room temperature. The samples were aliquoted into 100 μL eppendorf tubes and held using 1% agarose. T<sub>1</sub> relaxation times were measured either using the Fast-Imaging with Steady Precession (FISP) method<sup>[29]</sup> where TR/TE = 3.2/1.6 ms, flip angle = 60°, 40 τ<sub>1</sub> delay values of 40–3800 ms, or using a multi-point inversion recovery method using 8–16 τ<sub>1</sub> delay values of 17–8000 ms. T<sub>2</sub> relaxation times were measured using the Multi-Spin-Multi-Echo (MSME) method<sup>[30]</sup> employing a Carr-Purcell-Meiboom-Gill sequence (TR/TE = 5000/11 ms with 16 echos). The longitudinal (r<sub>1</sub>) and transverse (r<sub>2</sub>) relaxivities of a sample were determined by plotting the values of 1/T<sub>1</sub> and 1/T<sub>2</sub> versus the Gd(DTPA) millimolar concentration of a series of diluted suspensions of the same sample. In comparison, free [Gd(dtpa)]<sup>2-</sup> in solution was also measured in the relaxation times T<sub>1</sub> and T<sub>2</sub>, which resulted in r<sub>1</sub> and r<sub>2</sub>, respectively, under the same conditions (Table 2).

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- [1] P. Caravan, J. J. Ellison, T. J. McMurry, R. B. Lauffer, *Chem. Rev.* **1999**, *99*, 2293.
- [2] D. Delbecke, C. W. Pinson, *J. Hepatobiliary Pancreat. Surg.* **2004**, *11*, 4.
- [3] J. C. C. Moon, S. K. Prasad, *Curr. Cardiol. Rev.* **2005**, *7*, 39.
- [4] H. Degani, M. Chetrit-Dadiani, L. Bogin, E. Furman-Haran, *Thromb. Haemostasis* **2003**, *89*, 25.
- [5] R. Sanjay, P. Martin, *Cardiol. Clin.* **2002**, *20*, 501.
- [6] H. Kobayashi, M. W. Brechbiel, *Adv. Drug Delivery Rev.* **2005**, *57*, 2271.
- [7] E. Toth, L. Helm, A. E. Merbach in *Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging* (Ed.: A. E. T. Merbach), Wiley, Chichester, **2001**, p. 45.

- [8] S. Aime, M. Botta, M. Fasano, E. Terreno, *Acc. Chem. Res.* **1999**, *32*, 941.
- [9] J. L. Turner, D. Pan, R. Plummer, Z. Chen, A. K. Whittaker, K. L. Wooley, *Adv. Funct. Mater.* **2005**, *15*, 1248.
- [10] D. M. Doble, M. Botta, J. Wang, S. Aime, A. Barge, K. N. Raymound, *J. Am. Chem. Soc.* **2001**, *123*, 10758.
- [11] M. Woods, M. Botta, S. Avedano, J. Wang, A. D. Sherry, *Dalton Trans.* **2005**, 3829.
- [12] Z. Jaszberenyi, A. Sour, E. Toth, M. Benmelouka, A. E. Merbach, *Dalton Trans.* **2005**, 2713.
- [13] D. A. Fulton, M. O'Halloran, D. Parker, K. Senanayake, M. Botta, S. Aime, *Chem. Commun.* **2005**, 474.
- [14] R. Ruloff, E. Toth, R. Scopelliti, R. Tripier, H. Handel, A. E. Merbach, *Chem. Commun.* **2002**, 2630.
- [15] C. Platas-Iglesias, L. Vander Elst, W. Zhou, R. N. Muller, C. F. G. C. Geraldes, T. Maschmeyer, J. A. Peters, *Chem. Eur. J.* **2002**, *8*, 5121.
- [16] Y.-S. Lin, Y. Hung, J.-K. Su, R. Lee, C. Chang, M.-L. Lin, C.-Y. Mou, *J. Phys. Chem. B* **2004**, *108*, 15608.
- [17] G. A. Pereira, D. Ananias, J. Rocha, V. S. Amaral, R. N. Muller, L. Vander Elst, E. Toth, J. A. Peters, C. F. G. C. Geraldes, *J. Chem. Mater.* **2005**, *15*, 3832.
- [18] P. S. Braterman, Z. P. Xu, F. Yarberr in *Handbook of Layered Materials* (Eds.: S. M. Auerbach, K. A. Carrado, P. K. Dutta), Marcel Dekker, New York, **2004**, p. 373.
- [19] F. Cavani, F. Trifiro, A. Vaccari, *Catal. Today* **1991**, *11*, 173.
- [20] S. Y. Kwak, Y. J. Jeong, J. S. Park, J. H. Choy, *Solid State Ionics* **2002**, *151*, 229.
- [21] Z. P. Xu, G. Stevenson, C.-Q. Lu, G. Q. M. Lu, P. Bartlett, P. Gray, *J. Am. Chem. Soc.* **2006**, *128*, 36.
- [22] Z. P. Xu, P. S. Braterman in *Dekker Encyclopedia of Nanoscience and Nanotechnology* (Eds.: J. A. Schwars, C. I. Contescu, K. Putyera), Marcel Dekker, New York, **2004**, p. 3387.
- [23] W. W. Simons, *The Sadtler Handbook of Infrared Spectra*, Sadtler Research Laboratories, Pennsylvania, **1978**.
- [24] M. J. Hernandez-Moreno, M. A. Ulibarri, J. L. Rendon, C. J. Serna, *Phys. Chem. Miner.* **1985**, *12*, 34.
- [25] H. Gries, H. Miklantz, *Physiol. Chem. Phys. Med. NMR* **1984**, *16*, 105.
- [26] R. J. Hunter, *Zeta Potential in Colloid Science: Principles and Applications*, Academic Press, London, **1988**.
- [27] H. Cai, A. C. Hillier, K. R. Franklin, C. C. Nunn, M. D. Ward, *Science* **1994**, *266*, 1551.
- [28] Z. P. Xu, P. S. Braterman, *J. Mater. Chem.* **2003**, *13*, 268.
- [29] A. P. Crawley, R. M. Henkelman, *Magn. Reson. Med.* **1988**, *7*, 23.
- [30] J. D. De Certaines, W. M. M. J. Bovee, F. Podo, *Magnetic Resonance Spectroscopy in Biology and Medicine*, Pergamon, New York, **1992**.

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